Metagenomics: a flexible tool for discovery

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1. Metagenomics basics

2. Examples of using metagenomics

3. Integration with other approaches

What is metagenomics?

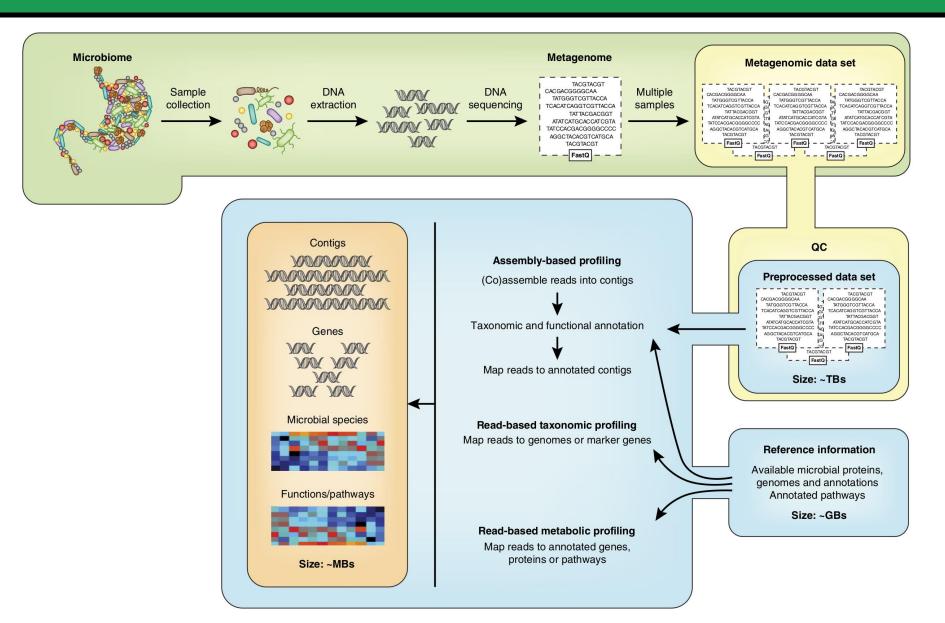


Metagenomics is the sequencing and analysis of DNA from whole communities (e.g. soils, waters, sediments, stool, built environment).

It provides a relatively unbiased view on the taxonomic composition and potential functions of communities (including bacteria, archaea, viruses).

This technology has rapidly expanded in the last decade through innovations in sequencing, assembly, and analysis.

A typical workflow for metagenome sequencing and analysis



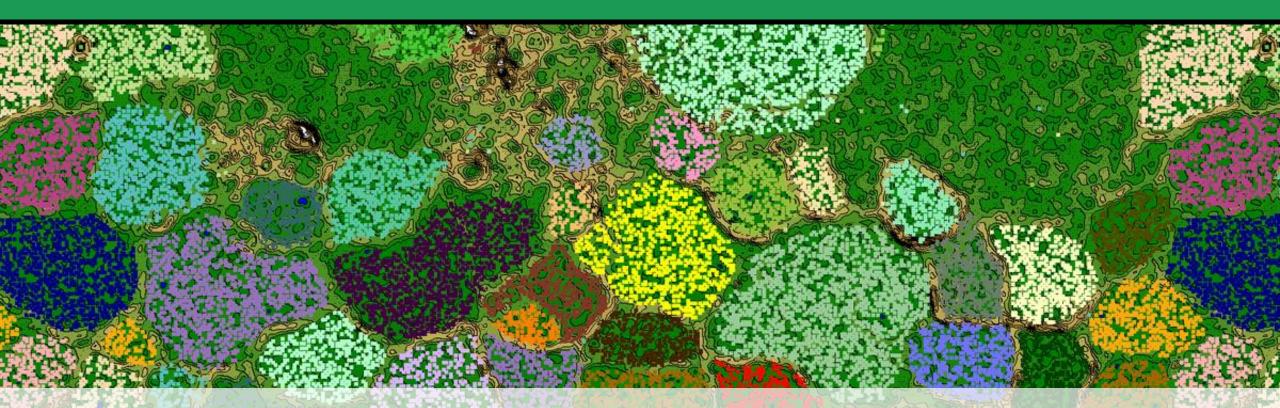
Two main types of sequencing available

	Short-read	Long-read
Platform	Illumina NextSeq, HiSeq, NovaSeq	Oxford Nanopore MiniION, PacBio Sequel
Method	Amplified templates sequenced by synthesis	Single molecule flow or synthesis in real-time
Output	Short paired-end reads (2 × 150 bp)	Long reads of variable length (1 kbp to 1 mbp)
Error rate	~0.1%	~10%
Cost	\$100/sample library prep, \$60/Gbp sequence	~\$250/Gbp library prep and sequencing



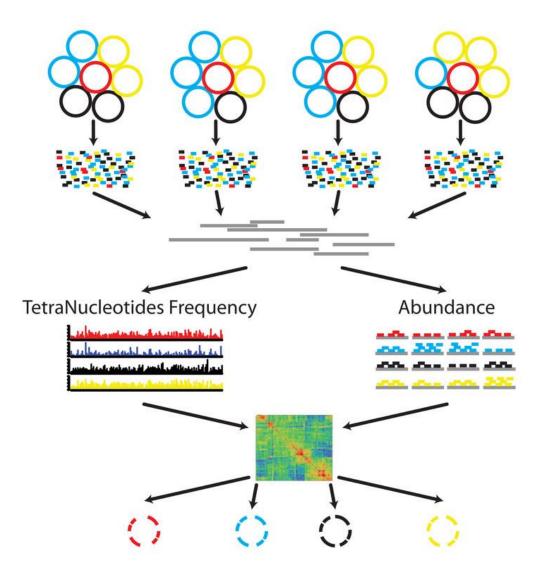


Gene-centric and genome-centric analyses



Software has been developed to assemble short and long reads into contigs. This enables gene-centric analyses. In turn, these contigs can be assembled into nearcomplete genomes. This enables genome-centric analyses.

Just one approach for assembly and binning



Sequencing and assembly

- 1. Samples from multiple sites or times
- 2. Metagenome libraries
- 3. Initial de novo individual or combined assembly

Binning with MetaBAT

- 4. Calculate TNF for each contig
- 5. Calculate abundance per library for each contig
- 6. Form and check genome bins iteratively

What is the composition and diversity of the community?

What are the functional capabilities of the community?

How might community members interact with the environment?

How do community members potentially interact with each other?

How does composition and function vary across time and space?

How can we use these findings to advance fundamental or scientific research?

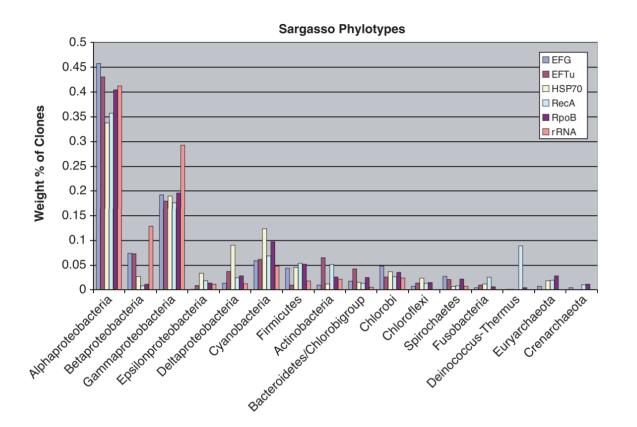
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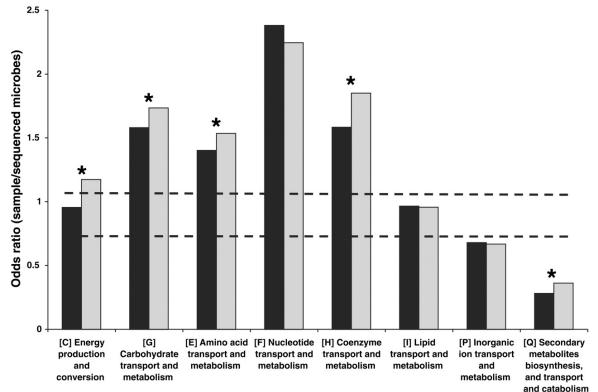
Venter et al., Science 2004

Metagenomic sequencing of the Sargasso sea detected 1800 phylotypes and 1.2M novel genes

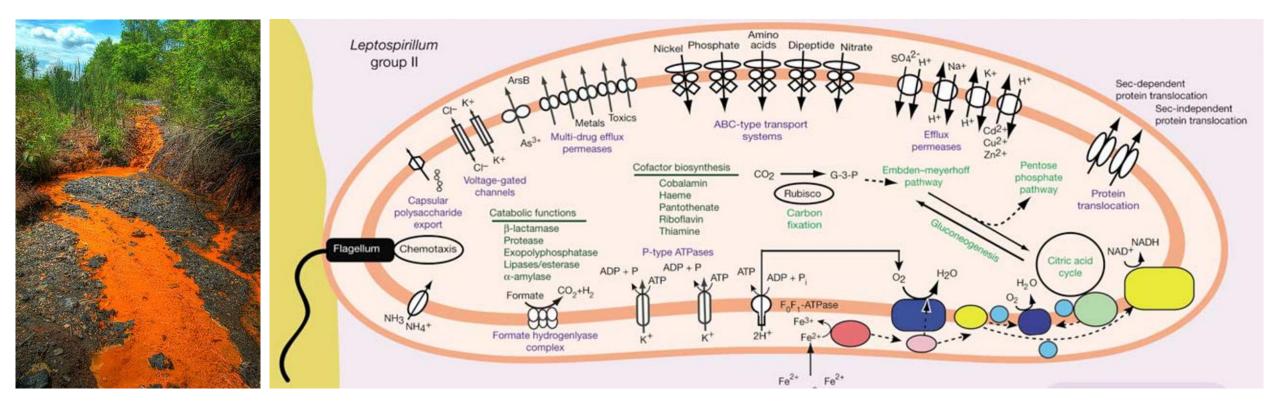


Gill et al., Science 2006

Metagenomic sequencing of the human gut revealed an enrichment of digestion genes



The first of many metagenome-assembled genomes



Unusual biology across a group comprising more than 15% of domain Bacteria

Christopher T. Brown¹, Laura A. Hug², Brian C. Thomas², Itai Sharon², Cindy J. Castelle², Andrea Singh², Michael J. Wilkins^{3,4}, Kelly C. Wrighton⁴, Kenneth H. Williams⁵ & Jillian F. Banfield^{2,5,6}

An environmental bacterial taxon with a large and distinct metabolic repertoire

Micheal C. Wilson^{1,2*}, Tetsushi Mori^{3*}, Christian Rückert⁴, Agustinus R. Uria^{1,2}, Maximilian J. Helf^{1,2}, Kentaro Takada⁵, Christine Gernert⁶, Ursula A. E. Steffens², Nina Heycke², Susanne Schmitt⁷, Christian Rinke⁸, Eric J. N. Helfrich^{1,2}, Alexander O. Brachmann¹, Cristian Gurgui², Toshiyuki Wakimoto⁹, Matthias Kracht², Max Crüsemann², Ute Hentschel⁶, Ikuro Abe⁹, Shigeki Matsunaga⁵, Jörn Kalinowski⁴, Haruko Takeyama³ & Jörn Piel^{1,2}

Methane metabolism in the archaeal phylum Bathyarchaeota revealed by genome-centric metagenomics

Paul N. Evans, $^{1\pm}$ Donovan H. Parks, $^{1\pm}$ Grayson L. Chadwick, 2 Steven J. Robbins, 1 Victoria J. Orphan, 2 Suzanne D. Golding, 3 Gene W. Tyson $^{1,4}\dagger$

Complex archaea that bridge the gap between prokaryotes and eukaryotes

Anja Spang¹*, Jimmy H. Saw¹*, Steffen L. Jørgensen²*, Katarzyna Zaremba-Niedzwiedzka¹*, Joran Martijn¹, Anders E. Lind¹, Roel van Eijk¹†, Christa Schleper^{2,3}, Lionel Guy^{1,4} & Thijs J. G. Ettema¹

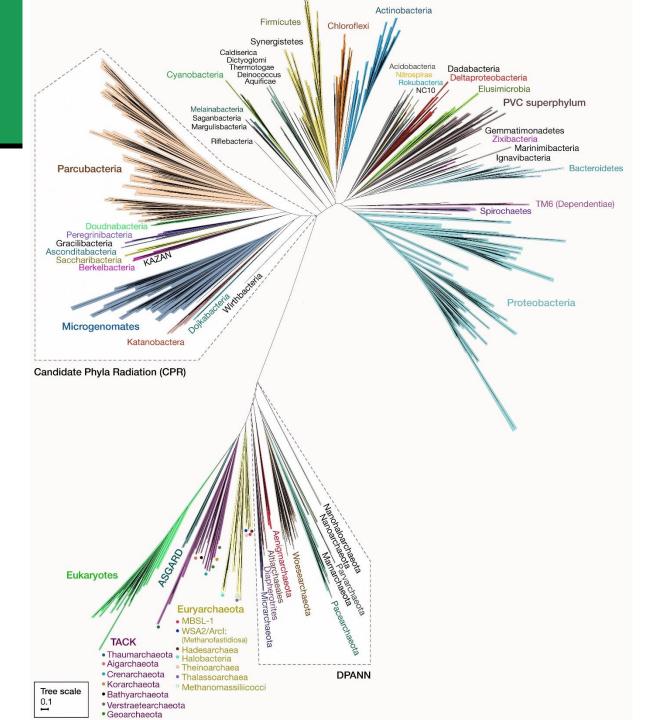
Atmospheric trace gases support primary production in Antarctic desert surface soil

Mukan Ji¹*, Chris Greening²*, Inka Vanwonterghem³, Carlo R. Carere⁴, Sean K. Bay², Jason A. Steen³, Kate Montgomery¹, Thomas Lines², John Beardall², Josie van Dorst¹, Ian Snape⁵, Matthew B. Stott⁴, Philip Hugenholtz³ & Belinda C. Ferrari¹

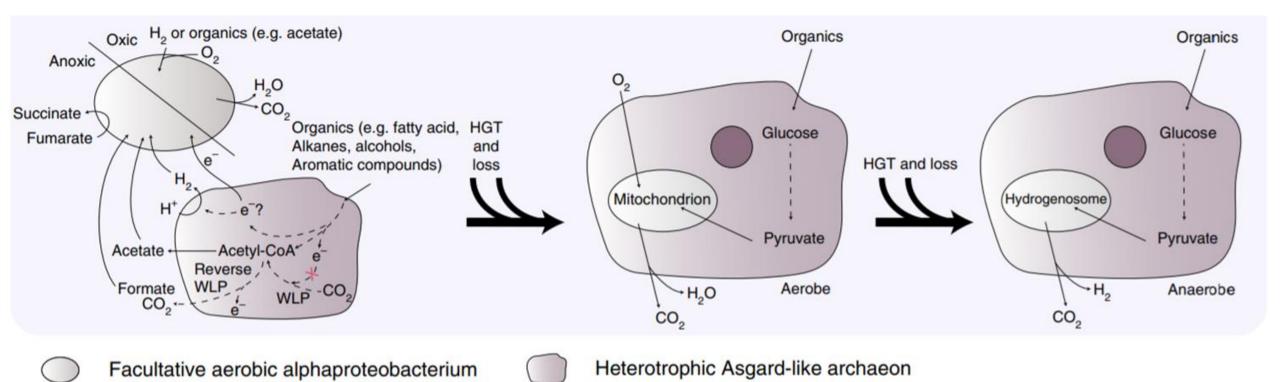
A highly abundant bacteriophage discovered in the unknown sequences of human faecal metagenomes

Bas E. Dutilh^{1,2,3,4}, Noriko Cassman^{3,†}, Katelyn McNair², Savannah E. Sanchez³, Genivaldo G.Z. Silva⁵, Lance Boling³, Jeremy J. Barr³, Daan R. Speth⁶, Victor Seguritan³, Ramy K. Aziz^{2,7}, Ben Felts⁸, Elizabeth A. Dinsdale^{3,5}, John L. Mokili³ & Robert A. Edwards^{2,4,5,9}

This has reshaped the tree of life...

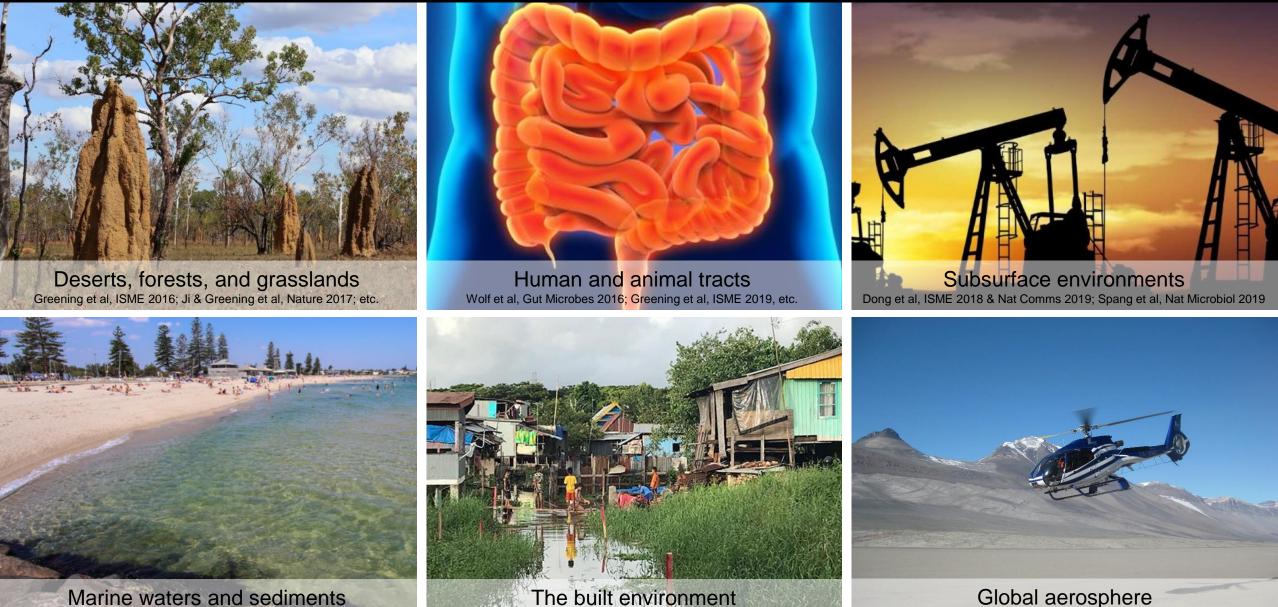


A recent study reconstructed the metabolism of the Asgard archaea. We proposed that these archaea were fermenters and their alphaproteobacterial partners recycled the endproducts. The bacteria eventually became mitochondria.



Spang et al (2019), Nat Microbiol 4, 1138-1148

We've used metagenomics to understand how various ecosystems function



Marine waters and sediments Kessleret al, Nat Microbiol 2019; Baltar et al., in preparation, etc.

The built environment Taruc et al., Lancet Planetary Health 2019; Lappan et al., in prep

Archer & Lappan et al., in prep

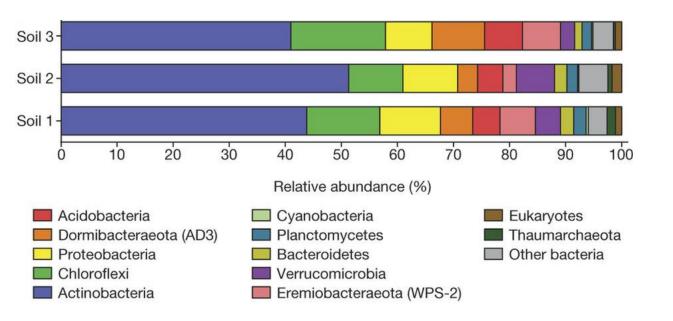
Some of these studies have changed the way we think about life



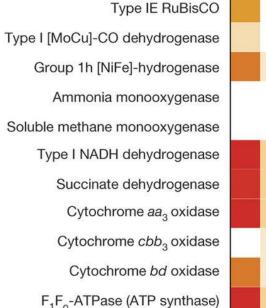
It's thought that most soils are sustained by sunlight. However, we provided strong evidence that atmospheric energy sources support the diverse, abundant communities in terrestrial Antarctica, namely hydrogen and carbon monoxide.

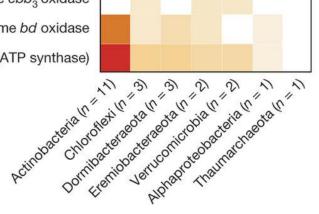
Genome-resolved metagenomics that was later validated

These soils are phylogenetically diverse but contain few phototrophs



Genome reconstructions show Antarctic soils dominated by gas scavengers



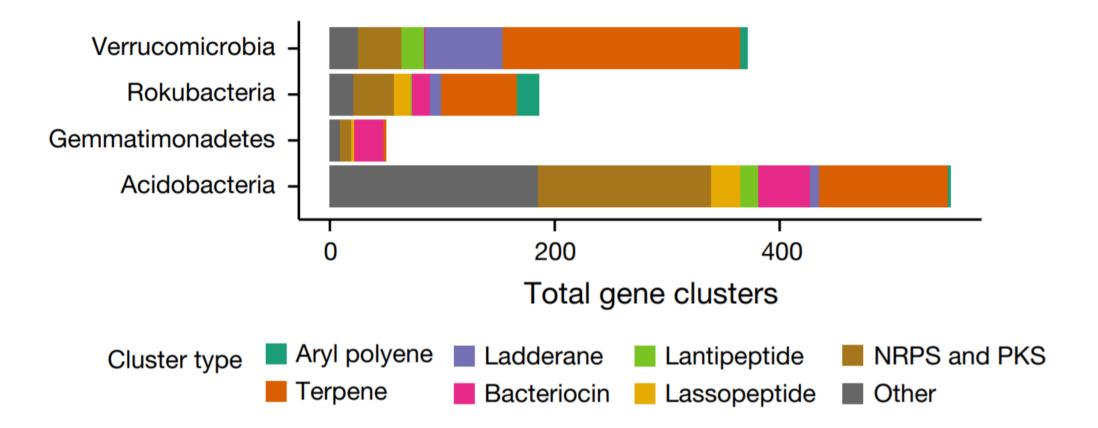


Metagenomics is being for diverse purposes: to profile the gastrointestinal microbiome, diagnose infectious disease, or track spread of antimicrobial resistance.





We are using metagenomics to profile antimicrobial resistance genes. We are looking at environmental samples, animal scats, and human stool before and after a public health intervention. For example, metagenomics has been used to discovery gene clusters for novel antibiotics across soil phyla such as the Acidobacteria and Verrucomicrobia.



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"Metagenomics can only provide inferences about how these organisms live, rather than any sort of proofs. Moreover, these approaches have limited capacity to discover or characterize genes encoding novel functions."

Going more "meta" can help to develop insights

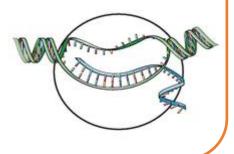
Metagenomics

- DNA sequencing and assembly.
- What is there?
- What can they do?



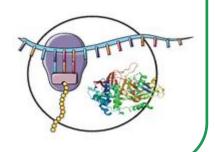
Metaproteomics

- Protein separation and mass spectrometry.
- What is translated?
- Are complexes formed?



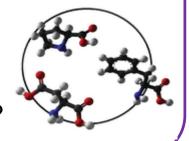
Metatranscriptomics

- RNA reverse transcription and sequencing.
- What is active?
- How do they respond?

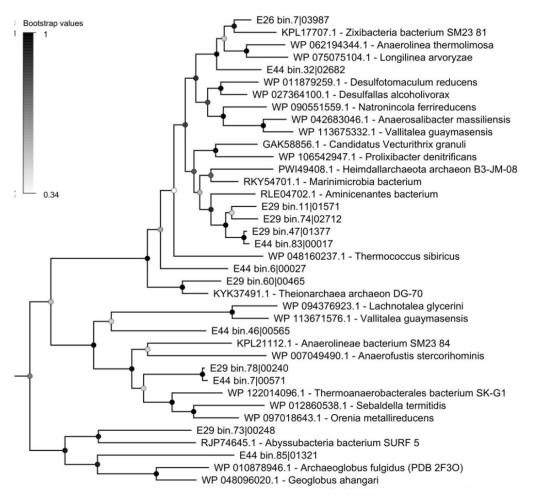


Metabolomics

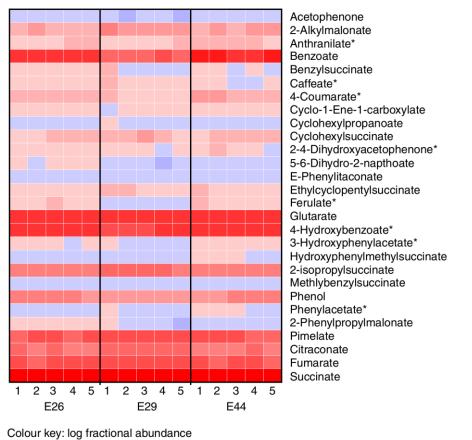
- Metabolite separation and mass spectrometry.
- What pathways are active?
- How is environment modified?

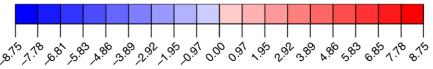


Many MAGs encode novel enzymes that may activate hydrocarbons by fumarate addition

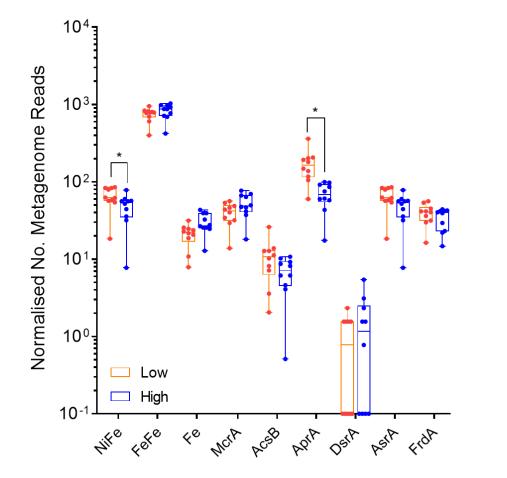


Metabolomics confirmed that succinatehydrocarbon conjugates are highly abundant

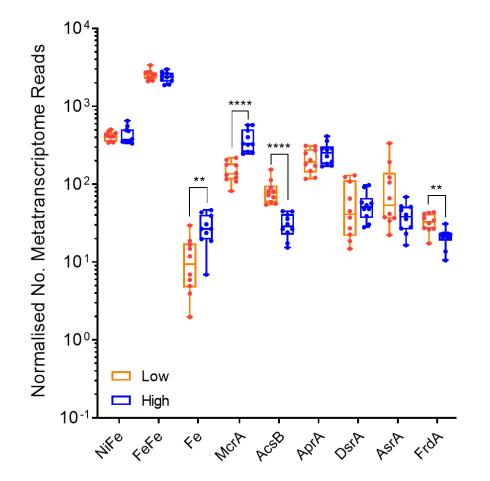




Few differences in community composition or gene abundance between low and high methane yield sheep



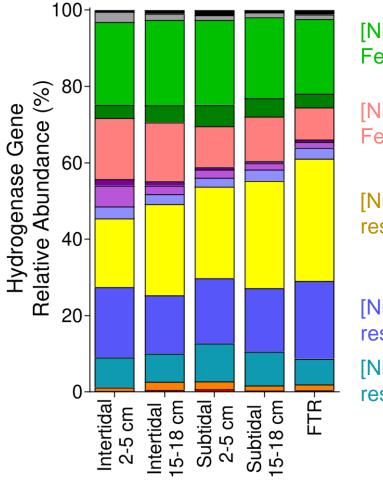
Methanogenesis downregulated in low yield sheep in favour of acetogenesis and fumarate reduction



"We must integrate meta-omics technologies with more targeted approaches. Only through such combinations will it be possible to gain both a broad and a deep understanding of how the uncultured majority live."

Linking microbial communities to biogeochemical function in beach sands

Metagenomic profiling suggests diverse capabilities for hydrogen metabolism



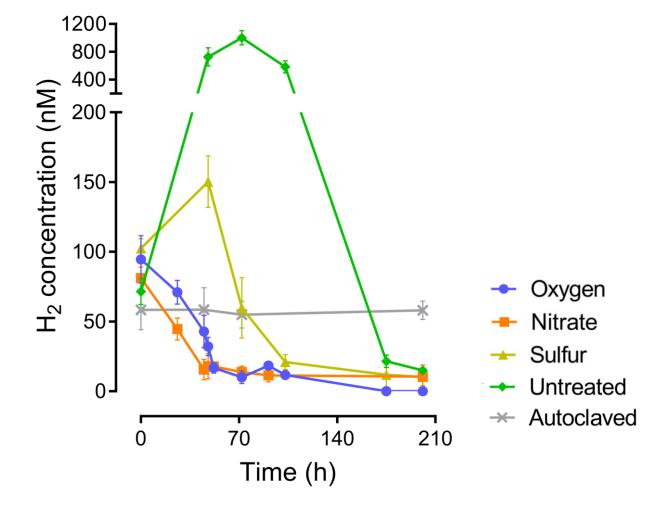
[NiFe] 3d: Fermentation

[NiFe] 3b: Fermentation

[NiFe] 1e: Sulfur respiration

[NiFe] 1d: Aerobic respiration [NiFe] 1c: Nitrate respiration

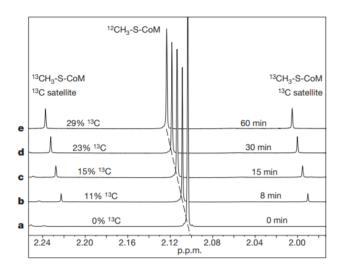
Biogeochemical perturbation studies perfectly validates metagenomic predictions



LETTERS

The key nickel enzyme of methanogenesis catalyses the anaerobic oxidation of methane

Silvan Scheller¹, Meike Goenrich², Reinhard Boecher², Rudolf K. Thauer² & Bernhard Jaun¹





Bacterial Rhodopsin: Evidence for a New Type of Phototrophy in the Sea

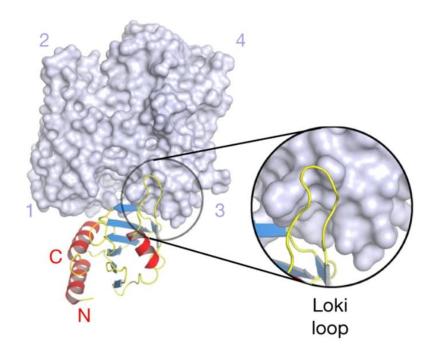
Oded Béjà,¹ L. Aravind,² Eugene V. Koonin,² Marcelino T. Suzuki,¹ Andrew Hadd,³ Linh P. Nguyen,³ Stevan B. Jovanovich,³ Christian M. Gates,³ Robert A. Feldman,³ John L. Spudich,⁴ Elena N. Spudich,⁴ Edward F. DeLong^{1*}



https://doi.org/10.1038/s41586-018-0548-6

Genomes of Asgard archaea encode profilins that regulate actin

Caner Akıl^{1,2} & Robert C. Robinson^{1,2,3}*

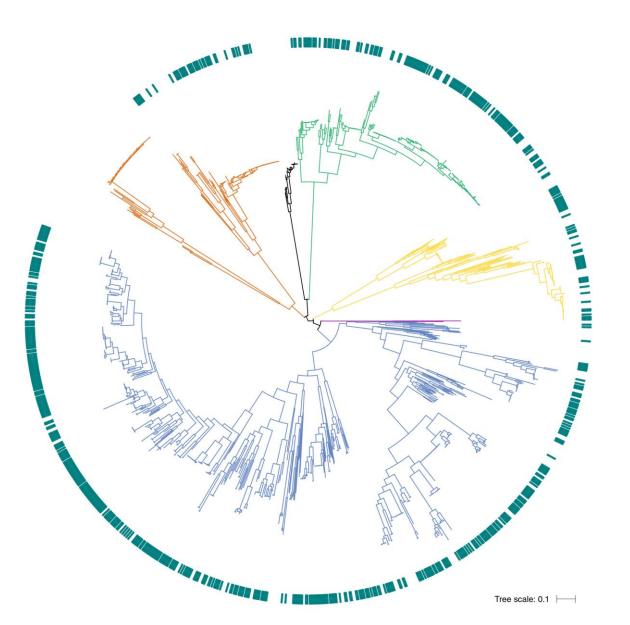


Going back to culture to bridge these divides

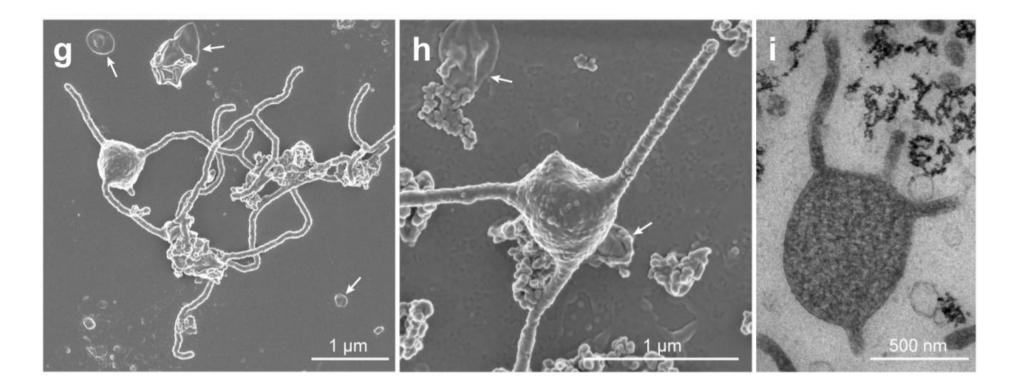
There has been a recent resurgence in culturing. Both targeted and untargeted approaches have been developed.

For example, researchers have performed high-throughput cultivation and sequencing of thousands of human gut bacterial isolates.

Researchers have now cultured organisms discovered by metagenomics, e.g. Lokiarchaeota, Atribacteria, Latesibacteria.



A spectacular example of this is the successful seven-year isolation of a Lokiarchaeum. This confirmed metagenomics-based inferences about its phylogeny, morphology, and metabolism, while providing many new discoveries.



Take-home messages

Metagenomics has become a standard technique to profile microbial communities. This has been driven by rapid developments in sequencing technologies and bioinformatics tools.

Metagenomics has provided unprecedented insights into the diversity of life, the functions of ecosystems, and various medical, environmental, and industrial questions.

However, it has become increasingly critical to test metagenomics inferences. It is possible to use meta-omic, culture-based, and biogeochemical approaches, among others, to do this.

Any questions? Thanks for listening!

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No more air for me, thanks





